THE INFLUENCE OF NITROGEN ON THE DECOMPOSITION OF CROP RESIDUES IN THE SOIL

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ABSTRACT

Additions of mineral nitrogen accelerated the initial decomposition rate of incorporated wheat straw, alfalfa hay and glucose when added to two soils differing widely in organic matter content. However, in the more advanced stages of decomposition the reverse was true, and over the total incubation period larger amounts of carbon were maintained in soils supplemented with nitrogen.

In contrast to all other residues used, nitrogen additions to cellulose effected a continuous and substantial increase in residue decomposition. This was the only residue for which the mineralization of soil organic matter did not supply nitrogen adequate for its decomposition within 120 days.

The very slow rate of decomposition of sphagnum peat could be attributed to its high lignin content, rather than to the nitrogen levels.

Sulphacetolysis analysis, which measures the non-humified carbon, indicated the feasibility of separating non-humified crop residues from the more complex soil organic matter. Addition of organic amendments thus resulted in a drop in the soil humification quotient. Nitrogen resulted in the retention of a significantly higher percentage of the added residue, without a drop in the humification quotient for the high organic matter Melfort soil.

Residue applications to soils produced a significant improvement of structural development, especially in the low organic matter soil (Arborfield).

INTRODUCTION

The long-term effect of nitrogen on the transformation of organic carbon within the soil, although of principal importance, is not clearly established. Since humus has been found to contain a fairly constant level of nitrogen under comparative environmental conditions (2, 12) it is usually argued that humus synthesis from crop residues, or other added organic amendments, cannot occur unless sufficient nitrogen is present. Inorganic nitrogen additions to a residue usually stimulate the initial rate of decomposition. This may lead to a higher degree of humification of the original plant remains, but not necessarily to a build-up of the soil organic matter level. Lee and Bray (3),Salter (10), and Turk and Millar (16) consider that added nitrogen is of principal importance for the prevention of excessive carbon loss. Other investigators utilizing different soils and environmental conditions have come to the opposite conclusion (8, 17).

There are also experimental results indicating insignificant differences in decomposition between nitrogen-treated and untreated residues in soil (4, 11). Newton (6), studying decomposition rates of incorporated wheat straw in Podzolic Grey Wooded, Chernozemic Black and Chernozemic Brown soils, did not obtain any significant responses in CO₂ evolution to fertilization with nitrogen.

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This study was conducted to investigate the problem of maintaining soil organic matter by the addition of various residue materials, with differing nitrogen levels, to two cultivated mineral soils of different organic matter content. Quantitative carbon mineralization studies, by means of measuring CO₂ production, and a qualitative characterization of the humus fraction, were used in an attempt to resolve the discrepancies among previously published reports.

MATERIALS AND METHODS

The two soils used in this study were obtained from the cultivated Ap horizon of an Orthic Black silty clay (Melfort Association), and a Grey Wooded Solodized Solonetz clay loam (Arborfield Association), respectively (5). Both soils were developed on fine textured, glacial-lacustrine parent material. They are both free of inorganic carbonate, but differ markedly in soil organic matter content. Some of the more important physical and chemical characteristics of the two soils are shown in Table 1.

Particle size distribution was determined by the pipette method, and pH values on a saturated paste. The organic carbon was determined by the dry combustion technique, and nitrogen by using the micro-Kjeldahl procedure.

Table 2 shows the composition of the five organic residues used in this study. Wheat straw, alfalfa hay and sphagnum moss were applied to the soil in a finely ground form. The cellulose was derived from shredded filter paper, and glucose was an analytical reagent. Total phosphorus in the residues was determined by the molybdo-phosphoric blue color method (14) after wet digestion of the material.

<table>
<thead>
<tr>
<th>Type of soil</th>
<th>Percentage¹</th>
<th>pH</th>
<th>Org. carbon %</th>
<th>Nitrogen %</th>
<th>C/N ratio</th>
<th>Hyg. moist. %</th>
<th>1/3 atmos. %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Melfort</td>
<td>18.7 Sand, 41.6 Silt, 39.7 Clay</td>
<td>6.40</td>
<td>6.00</td>
<td>0.491</td>
<td>12.22</td>
<td>4.40</td>
<td>38</td>
</tr>
<tr>
<td>Arborfield</td>
<td>37.0 Sand, 38.0 Silt, 25.0 Clay</td>
<td>5.85</td>
<td>1.64</td>
<td>0.131</td>
<td>12.52</td>
<td>1.24</td>
<td>23</td>
</tr>
</tbody>
</table>

¹Sand: 2 mm. to 0.05 mm.
Silt: 0.05 mm. to 0.002 mm.
Clay: finer than 0.002 mm.

<table>
<thead>
<tr>
<th>Organic compound</th>
<th>%C</th>
<th>%N</th>
<th>%P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wheat straw</td>
<td>47.1</td>
<td>0.787</td>
<td>0.078</td>
</tr>
<tr>
<td>Alfalfa</td>
<td>45.3</td>
<td>2.378</td>
<td>0.134</td>
</tr>
<tr>
<td>Sphagnum peat</td>
<td>45.3</td>
<td>0.713</td>
<td>0.050</td>
</tr>
<tr>
<td>Cellulose (filter paper)</td>
<td>41.0</td>
<td>none</td>
<td>none</td>
</tr>
<tr>
<td>Glucose (pure chemical)</td>
<td>40.0</td>
<td>none</td>
<td>none</td>
</tr>
</tbody>
</table>
Fifty grams of soil, plus residue applied at a rate corresponding to 4, 20 or 40 tons per acre (0.4, 2 and 4 per cent respectively) were incubated in 500-milliliter Erlenmeyer flasks. All residues were adjusted to a uniform carbon-phosphorus ratio (250 to 1) by the addition of K\textsubscript{2}HPO\textsubscript{4}. The carbon to nitrogen ratios of the residues were adjusted by the addition of NH\textsubscript{4}NO\textsubscript{3}. The rate of decay of the incorporated residue was followed by measuring the evolved CO\textsubscript{2} using an aeration train; the CO\textsubscript{2} was collected in 0.5 N NaOH scrubbers filled with "Berl Saddles" to increase the absorbing surface. All CO\textsubscript{2} values obtained from the incorporated residues were adjusted for CO\textsubscript{2} evolved from an untreated check soil during the same period. The moisture content of the samples was maintained at approximately 70 per cent of the field capacity and the samples were incubated at room temperature.

The organic matter in some of the incubated treatments was fractionated by Springer's sulphacetolysis technique (13, 15). The extracting solvent consisted of a mixture of glacial acetic acid, acetic anhydride, and sulphuric acid. This treatment dissolves certain organic matter fractions which are characterized by a low degree of polymerization, and low nitrogen content. These soluble fractions of organic matter consist chiefly of humin lignin acids and fulvic acids, while the insoluble part contains humic acids.

**Figure 1.** Cumulative CO\textsubscript{2} evolution from untreated soils.
The soil organic matter was further characterized following, in principle, the procedure as described by Reissig (9). This procedure did not indicate any differences in organic matter composition attributable to crop residues.

RESULTS

The respiratory activity of the unamended soil samples is shown in Figure 1. A comparison of the curves indicates slightly higher CO$_2$ evaluation from the Melfort soil sample than from the Arborfield soil. When expressed as a per cent of soil organic matter mineralized, however, the Melfort soil sample lost 2.74 per cent and the Arborfield 7.95 per cent of its organic matter during the 150-day incubation period.

The effect of nitrogen fertilization on the evolution of CO$_2$ from the high organic matter Melfort soil, and of calcium on the evolution from the slightly acidic Arborfield soil, was also determined. The addition of 0.078 per cent nitrogen to the Melfort soil, without added residue, resulted in a 63 per cent increase in CO$_2$ evolved during a 32-day incubation period. Similarly, raising the pH of the Arborfield soil sample to approximately neutral by the addition of calcium hydroxide resulted in an enhanced de-
composition rate. Sixty-five milligrams of calcium hydroxide increased the total amount of CO₂ evolved from 153 to 274 milligrams.

**Decomposition of Straw and Alfalfa**

Figure 2 illustrates the cumulative CO₂ evolution obtained from the two soils treated with wheat straw at a rate equivalent to 4 tons per acre. Alteration of the original carbon to nitrogen ratio of the straw from 60/1 to 10/1 by the addition of nitrogen resulted in a reduced CO₂ production during the 114-day incubation period. After 2½ months, the rate of mineralization of the straw was practically zero in both soils. Approximately 20 per cent of the added residue was not accounted for by the CO₂ evolved and is assumed to be a contribution to soil organic matter.

Figure 3 shows the cumulative CO₂ production curves for the two soils treated with a straw application equivalent to 40 tons per acre. The nitrogen-treated samples had higher initial decay rates during the first 3 weeks of incubation but lower rates thereafter. Nitrogen fertilization again resulted in a net reduction in total CO₂ evolved during the incubation period. The secondary CO₂ evolution peak observed during the 60- to 100-day period in the soil-straw mixture in the Arborfield soil was accompanied by intensive growth of fungal mycelia. Additional experiments with 20 tons of straw per acre adjusted to carbon to nitrogen ratios varying from 10/1 to 60/1 produced comparable results.

![Figure 3](https://example.com/figure3.png)

**Figure 3.** Effect of nitrogen on cumulative CO₂ evolution from soils incubated with 40 tons of wheat straw per acre. *Between arrows:* abundant fungi mycelia growth in Arborfield soil mixture.
Figures 4 and 5 represent cumulative CO$_2$ evolution curves obtained from the two soils treated with the equivalent of 4 and 40 tons per acre, respectively, of finely ground alfalfa hay having a C/N ratio of 19/1. At the lower level of residue addition, no effect due to the added nitrogen was observed for the Melfort soil; the Arborfield soil, however, which is low in organic matter, had a net decrease of CO$_2$ evolution. The decomposition reactions occurring in the 4-tons-per-acre-of-alfalfa treatment were practically completed in both soils by the end of 114 days with approximately 70 per cent of the carbon of the residue evolved as CO$_2$.

A comparison between the 40-tons-per-acre application of alfalfa and wheat straw indicates that alfalfa had a much higher initial rate of decay than the straw. This, however, was reversed in the more advanced stages of decomposition. Where the straw was not supplemented with nitrogen, CO$_2$ was still being evolved at a substantial rate, even after 150 days. In the comparable alfalfa treatments the evolution rate had, however, levelled off after 30 days’ incubation.
Figure 5. Effect of nitrogen on cumulative CO₂ evolution from soils incubated with 40 tons of alfalfa per acre.

Decomposition of Sphagnum Peat, Cellulose and Glucose

Sphagnum peat incorporated into the soil was decomposed at an extremely slow rate (Figure 6). With an application equivalent to 40 tons per acre, only 3 per cent and 4 per cent was decomposed during 5 months in the Arborfield and Melfort soils, respectively. Nitrogen additions to reduce the natural C/N ratio of the peat from 60/1 down to 10/1 lowered this rate further. The peat had a pH of 3.5, and reduced the pH of both soils by approximately 0.7 units. The addition of ammonium nitrate further increased the acidity so that the Melfort soil dropped from an original pH value of 6.4 down to 4.8. The increased acidity did not, however, inhibit
mineralization of the residue for neutralization of both soils with calcium hydroxide resulted in a slight flush of activity which tended to level off after 14 days (Figure 6).

The depression of evolution of CO₂ by the addition of nitrogen to alfalfa, wheat straw, and peat residues in the soil indicated that the addition of nitrogen should lead to the formation of increased amounts of soil humus. Since this observation is somewhat contentious, the effect of nitrogen on the decomposition of cellulose and glucose was also investigated. Cellulose adjusted to a carbon to nitrogen ratio of 10/1 and, incorporated in both soils, had an initial decomposition rate almost double that of the soil-cellulose mixture without added nitrogen (Figure 7). The reversal in decay rates due to nitrogen, which was observed for almost all other residue treatments, did not however occur during 114 days of incubation.

The addition of nitrogen with glucose, which is also a nitrogen-free compound, resulted in decomposition trends opposite to those observed with cellulose and in agreement with those for straw and alfalfa (Figures 2, 3, 4, 5). The added nitrogen only caused a significant increase in the rate of CO₂ evolution during the first few days of incubation. Thus, when nitrogen was supplied to the Melfort soil with glucose, approximately 60 per cent of the glucose carbon was oxidized within the first 4 days of incubation. Nevertheless, the nitrogen-treated samples had a lower net evolution of CO₂.
### Table 3. — Incorporated residues remaining after completion of the incubation period, and state of humification of the applied residue

<table>
<thead>
<tr>
<th>Residue and rates of application tons/acre</th>
<th>C/N ratio of residues</th>
<th>Residue remaining %</th>
<th>Balance due to N, tons/acre</th>
<th>Melfort soil</th>
<th>Arborfield soil</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Total carbon in % C_T</td>
<td>Humus(^a) carbon in % C_H</td>
<td>Humification quotient C_H / C_T \times 100</td>
</tr>
<tr>
<td>Soil alone</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Straw — 4</td>
<td>60/1</td>
<td>18</td>
<td>6.08</td>
<td>4.31</td>
<td>70.9</td>
</tr>
<tr>
<td>Straw — 4</td>
<td>10/1</td>
<td>23 +0.2</td>
<td>6.13</td>
<td>4.30</td>
<td>70.2</td>
</tr>
<tr>
<td>Straw — 40</td>
<td>60/1</td>
<td>35 +7.2</td>
<td>6.74</td>
<td>4.16</td>
<td>66.2</td>
</tr>
<tr>
<td>Straw — 40</td>
<td>10/1</td>
<td>53 +7.2</td>
<td>6.86</td>
<td>4.56</td>
<td>66.5</td>
</tr>
<tr>
<td>Alfalfa — 4</td>
<td>19/1</td>
<td>31</td>
<td>6.09</td>
<td>4.32</td>
<td>71.0</td>
</tr>
<tr>
<td>Alfalfa — 4</td>
<td>10/1</td>
<td>31 0</td>
<td>6.08</td>
<td>4.30</td>
<td>71.6</td>
</tr>
<tr>
<td>Alfalfa — 40</td>
<td>19/1</td>
<td>37 +2.8</td>
<td>6.63</td>
<td>4.27</td>
<td>64.4</td>
</tr>
<tr>
<td>Alfalfa — 40</td>
<td>10/1</td>
<td>44 +2.8</td>
<td>6.78</td>
<td>4.58</td>
<td>64.7</td>
</tr>
<tr>
<td>Peat — 40</td>
<td>63/1</td>
<td>96 +0.6</td>
<td>7.45</td>
<td>4.68</td>
<td>62.9</td>
</tr>
<tr>
<td>Peat — 40</td>
<td>10/1</td>
<td>98 +0.6</td>
<td>7.55</td>
<td>4.67</td>
<td>61.8</td>
</tr>
<tr>
<td>Cellulose — 40</td>
<td>no N.</td>
<td>64</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Cellulose — 40</td>
<td>10/1</td>
<td>39 −10.0</td>
<td>—</td>
<td>—</td>
<td>−</td>
</tr>
<tr>
<td>Glucose — 40</td>
<td>no N.</td>
<td>−3</td>
<td>5.97</td>
<td>3.99</td>
<td>66.8</td>
</tr>
<tr>
<td>Glucose — 40</td>
<td>10/1</td>
<td>11 +5.6</td>
<td>6.28</td>
<td>4.22</td>
<td>67.2</td>
</tr>
</tbody>
</table>

\(^a\)CO\(_2\) evolved minus CO\(_2\) from soil alone

\(^a\)Humus carbon = carbon left after extraction
for, in the nitrogen-treated samples, the rates of mineralization decreased to a markedly low level after the initial flush of activity during the first few days. The Melfort soil plus glucose, without supplemental nitrogen, demonstrated a net loss of organic carbon, whereas the addition of nitrogen decreased the CO₂ evolution 15 per cent (equivalent to 4.4 tons of glucose per acre).

An organic matter fractionation of the various incubated soils was carried out on samples after 100 days of incubation. According to the principle of Springer's sulphacetolysis separation (13, 15), non-humified and partially humified materials are separated and thus go into solution. The humus fractions, which are not as reactive, are not dissolved. The humification quotient \( \frac{\text{non-extractable carbon (Cₙ)}}{\text{total organic carbon (Cₒ)}} \times 100 \) is a relative measure of the stable soil humus. The results in Table 3 indicate that the humification quotients of the Arborfield soil samples for both the untreated and treated soils were lower than those obtained for the corresponding treatments in the Melfort soil. This is in agreement with the generally accepted view that the Black soils (Melfort) contain a higher ratio of more stable humus compounds than do the Grey (Arborfield) soils.

Residue application, especially at the higher rates of application, resulted in a marked lowering of the humification quotient. This was especially true for the peat treatments which were resistant to microbial attack to the extent that 97 per cent of residue remained after 154 days' incubation. The peat residue was, however, removed by the sulphacetolysis treatment and, although resistant to microbial attack, could thus not be considered to be humified.

Nitrogen additions to the glucose- and straw-treated samples resulted in the retention of an extensive amount of residue. The humification quotient of the Arborfield soil dropped as a result of the increased residue. In the Melfort soil, however, although as much as 7 tons of residue were retained due to the added nitrogen, the humification quotient was identical to the non-nitrogen-treated sample.

**Structural Development of Two Soils under the Influence of Various Residue Applications and Carbon to Nitrogen Ratios**

Treatments as used for the 5-month CO₂ evolution test were duplicated for both soils in a parallel set-up in order to permit some chemical and physical analyses, without disturbing the samples on which the CO₂ measurements were being made. These latter soils were incubated in Neubauer dishes, and the moisture content was kept at 70 per cent of the field capacity. Mechanical stirring was carried out once a month.

The soils developed characteristic structures during the 5-month period, indicating a distinct relationship with the applied treatments. This was particularly apparent for the Arborfield soil, which is characterized by weakly developed structure and tends to puddle when water is applied. Figure 8 shows the structural development of Arborfield soils incubated with glucose, straw and cellulose residues, with additional nitrogen in some cases.
The higher treatments were fully effective in diminishing the surface sealing, and in producing aggregates which withstood the dispersing action of applied water.

The type of residue influenced structural development to different degrees. Alfalfa produced a more friable structure than straw. Peat created a spongy structure in both soils (not shown), but no crumb formation was observed and surface sealing was aggravated. Glucose tended to cause the formation of hard non-friable aggregates, particularly in the Arborfield soil.

The addition of nitrogen had a favorable influence on structural development of most of the residue-soil mixtures. Although surface sealing appeared to be aggravated, the nitrogen treatment produced a finer aggregation and a greater friability in the straw, cellulose and glucose treatments. The alfalfa and peat treatments were not affected by nitrogen additions.

**DISCUSSION AND SUMMARY**

Results from this study indicate that added nitrogen affected residue decomposition and organic matter accumulation in two distinct ways. Nitrogen accelerated the initial decay rate of incorporated residues. This trend was, however, later reversed so that larger amounts of organic carbon were maintained when supplemental nitrogen was applied. This was true for all residues, except cellulose, which showed a consistently higher rate of decomposition when nitrogen was added. This suggests that a substantial nitrogen deficiency occurred only in the cellulose-treated samples. In all other treatments, after the initial flush of activity due to the added nitrogen, the second effect of nitrogen predominated, namely the formation of humified organic matter. Andrews (1) observed a similar phenomenon. He obtained high positive correlation coefficients between the nitrogen content of rye straw and the rates of CO₂ evolution during the first month of incubation. Thereafter, the correlation coefficient became negative and was as high as -0.853 at the end of the fourth month of incubation. The total carbon loss was, nevertheless, higher in the high nitrogen samples. It must be considered, however, that, in the experiment conducted by Andrews, only about one-half of the incorporated residue had been decomposed at the end of the experiment. The total effect of nitrogen on residue decomposition can thus only be measured by carefully controlled experiments extending over a long incubation period.

The decomposition of glucose was most advanced at all times during the decomposition period. Differences in total glucose decomposed due to nitrogen were quite apparent for the Melfort soil, but were just beginning to be discernible in the Arborfield soil. For the Melfort soil, a net gain equivalent to 4.4 tons of organic matter per acre was derived from the glucose residue with supplemental nitrogen. This compared very favorably with a net loss of 1.2 tons in the non-nitrogen-treated sample.

The humus fractionation tests disclosed the interesting fact that the sulphaetolysis extraction technique does differentiate between residue carbon and the more polymerized humus fraction. In the Melfort soil the
Figure 8. Structural development of Adirondack soil due to residue and nitrogen incorporation.

Top row: (no nitrogen)
- Check
- Glucose at 40 tons/acre
- Straw at 40 tons/acre
- Cellulose at 40 tons/acre

Bottom row: (10% residue
- Check
- Glucose at 40 tons/acre
- Straw at 40 tons/acre
- Cellulose at 40 tons/acre
- C/N of residue = 10/1
addition of nitrogen, although resulting in a substantial saving of organic carbon, did not result in a lowering of the humification quotient. This was not apparent in the Arborfield soil, perhaps due to the less advanced degree of decomposition of the incorporated residues.

The initially available nitrogen content of the soil, the mineral composition of the residues, and their major organic constituents, such as carbohydrates, proteins, lignins, fats and waxes will all affect the decomposition characteristics. Thus, the extremely low rates of mineralization of sphagnum peat residues, whether incubated with or without nitrogen, were due to the composition of the residue rather than to soil type or amendment.

The two soils utilized in this study differed markedly in physical and chemical characteristics. No fundamental differences were observed in the general character of the decomposition curves. The respiration of both soils was approximately equivalent in magnitude. However, it is noteworthy that there was a very significant difference in the structural changes between the two soils.

Although added nitrogen initially stimulated decomposition, there was no evidence (except for pure cellulose) that a lack of nitrogen inhibited the breakdown of the added residues. Under field conditions in Western Canada, mineralization of soil organic matter, especially in a summerfallow rotation, probably supplies adequate nutrients for residue decomposition and for growth of the following crop. The advantages to be gained by adding nitrogen to lower the losses of carbon from the added residues would not justify the expense, unless the crops respond to the added nutrients.

REFERENCES


